

Raucaffrinoline

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Key indicators

Single-crystal X-ray study
 $T = 297$ K
Mean $\sigma(\text{C}-\text{C}) = 0.005$ Å
 R factor = 0.040
 wR factor = 0.107
Data-to-parameter ratio = 7.6For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

In the crystal structure of the title compound [systematic name: (17*R*,20*α*,21*β*)-1,2-didehydro-1-demethyl-19-hydroxy-21-methyl-18-norajmalan-17-yl acetate], $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_3$, an intermolecular $\text{O}-\text{H}\cdots\text{O}$ hydrogen bond [2.810 (4) Å and 160°] stabilizes the molecular packing as an infinite chain along the [100] direction.

Comment

Bioactive indole alkaloids have been isolated from plant species of the Apocynaceae family, including many belonging to the *Rauvolfia* genus, among which *Rauvolfia serpentina* Benth, from India, has been an important source of reserpine (Stöckigt, 1995). The chemistry of the *Rauvolfia* species has been exhaustively investigated for the presence of indole alkaloids (Siddiqui *et al.*, 1987; Stöckigt, 1995; Court, 1983; Libot *et al.*, 1986). In a research programme on the occurrence of indole alkaloids in Brazilian species of *Rauvolfia*, a phytochemical analysis of the extracts from leaves and bark of *Rauvolfia bahiensis* A. DC. is described.

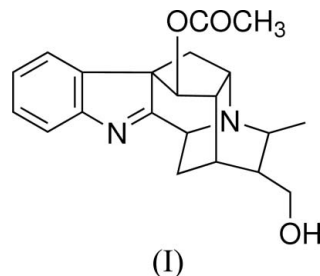


Fig. 1 shows the molecular structure of the title compound, (I), with the atomic numbering. The displacement ellipsoids of atoms O1 and O3 are highly anisotropic, indicating probable unresolved disorder in these sites.

The molecule contains an indole unit fused to a quinuclidine unit, forming a cage which can be described by five-, six- and seven-membered rings. The C8–C12 ring has puckering parameters (PP) (Cremer & Pople, 1975) $Q = 0.494$ (4) Å and $\varphi = 199.1$ (5)°, showing a distorted half-chair conformation. The rings N2/C10/C11/C15/C14/C13, N2/C17/C16/C15/C11/C10 and N2/C13/C14/C15/C16/C17 have PP Q (Å), θ (°) and φ (°) averaging 0.858 (4), 88.8 (3) and 160.9 (3), respectively, showing a twist–boat conformation. The ring N2/C10/C9/C8/C7/C13 has PP 0.655 (4), 19.9 (4) and 102.2 (10), respectively, showing a distorted chair conformation. The seven-membered ring C7/C8/C12/C11/C15/C14/C13 has PP $Q_2 = 0.703$ (4) Å, $Q_3 = 0.704$ (4) Å, $Q = 0.995$ (4) Å, $\varphi_2 = 215.5$ (3)° and $\varphi_3 = 11.6$ (3)°, showing a distorted twist–chair conformation.

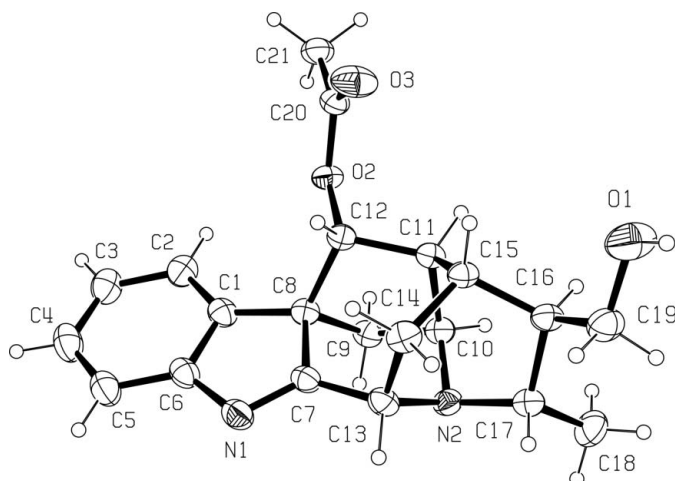


Figure 1
A view of (I) with the atom-numbering scheme. Displacement ellipsoids are drawn at the 30% probability level and H atoms are shown as small spheres of arbitrary radii.

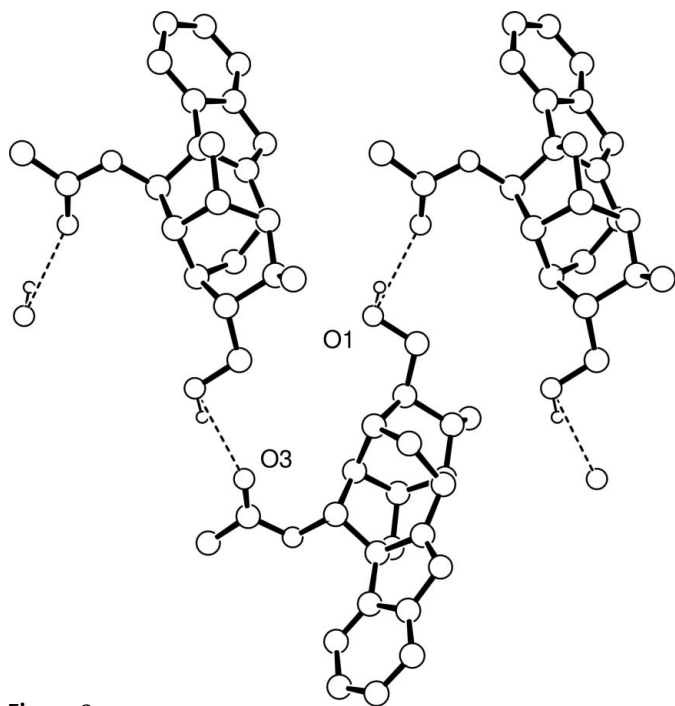


Figure 2
A packing diagram, viewed down the *b* axis; the *a* axis points to the right. Intermolecular O—H...O hydrogen bonds are shown as dashed lines. Only H atoms involved in hydrogen bonds are shown.

The bonds C7—C13, C16—C19 and C20—C21 are shortened by 0.027, 0.029 and 0.05 Å, respectively, compared with the expected formal bond lengths. The shortening of the C7—C13 and C20—C21 bonds might be ascribed to hyperconjugation. The C7—N1 bond length indicates a double bond and the angles around C7 confirm *sp*² hybridization. We have found two systems of fused rings similar to (I) in the Cambridge Structural Database (CSD, Version 5.27; Allen, 2002), with refcodes JAKFAC (C₃₀H₃₄N₂O₆; Li & Cook, 1998) and AJMALN10 (C₂₀H₂₆N₂O₂·CH₂Cl₂; Prewo & Stezowski, 1978). The superposition of these structures on that of (I) shows that the average r.m.s. deviation of the eight atoms of

the quinuclidine unit is 0.036 Å. The N2—C17—C16—C15 torsion angle (AJMALN10 14.57°; JAKFAC 9.59°) changes due to steric effects caused by different functional groups at positions C17 and C16. Selected geometric parameters are shown in Table 1.

The molecular packing of (I) is stabilized through a hydrogen-bonding network, shown in Fig. 2, with geometric parameters in Table 2. An intermolecular O1—H1...O3ⁱ hydrogen bond links neighbouring molecules related by the screw axis, resulting in an infinite one-dimensional chain along the [100] direction.

Experimental

Material from *Rauvolfia bahiensis* A. DC. was collected from Barra do Una, Ilhéus city (Bahia, Brazil), and the aerial parts were air-dried, powdered and extracted with 95% aqueous EtOH. The ethanolic extract from the leaves was submitted to acid/base treatment and fractions corresponding to different pH ranges were purified by repeated separation by preparative thin-layer chromatography, leading to the isolation of thirteen alkaloids, the known alkaloids raucaffrinoline, picrinine, vinorine, normacusine B, norseredamine, seredamine, 10-methoxynormacusine B, norpurpeline, purpeline, 12-methoxy-*N*-methyl-vellosimine, demethoxy-purpeline, 12-methoxyaffinisine and 12-methoxy-vellosimine (Kato *et al.*, 2002). The alkaloid raucaffrinoline, (I), was isolated as a brown powder. The spectroscopic data (NMR) and the melting point of raucaffrinoline are in complete accordance with those reported in the literature (Djerassi *et al.*, 1966; Solis *et al.*, 1993). Crystals of (I) suitable for single-crystal X-ray diffraction studies were obtained as brown blocks by recrystallization from methanol and hexane (1:9) (m.p. 522–524 K; Libot *et al.*, 1980). Spectroscopic analysis: ¹H NMR (CDCl₃/TMS 0.01%, 300 MHz, δ, p.p.m.): 7.47 (*d*, 1 H, *J* = 7 Hz, H2), 7.22 (*t*, 1 H, *J* = 7 Hz, H3), 7.38 (*t*, 1 H, *J* = 7 Hz, H4), 7.60 (*d*, 1 H, *J* = 7 Hz, H5), 2.80 (*dd*, 1 H, *J* = 12 and 5 Hz, H9A), 1.63 (*dd*, 1 H, *J* = 12 Hz, H9B), 3.65 (*m*, 1 H, H10), 2.38 (*t*, 1 H, *J* = 6 Hz, H11), 5.00 (*s*, 1 H, H12), 4.15 (*d*, 1 H, *J* = 10 Hz, H13), 1.94 (*dd*, 1 H, *J* = 15 and 10 Hz, H14A), 1.54 (*dd*, 1 H, *J* = 15 and 6 Hz, H14B), 2.48 (*m*, 1 H, H15), 1.54 (*dd*, 1 H, *J* = 15 and 6 Hz, H16), 2.52 (*m*, 1 H, H17), 1.27 (*d*, 3 H, *J* = 7 Hz, H18), 3.75 (*dd*, 2 H, *J* = 11 and 6 Hz, H19), 2.17 (*s*, 3 H, H21); ¹³C NMR (CDCl₃/TMS 0.01%, 75.0 MHz, δ, p.p.m.): 136.40 (C1), 123.8 (C2), 125.4 (C3), 128.6 (C4), 120.9 (C5), 156.5 (C6), 183.5 (C7), 65.0 (C8), 37.5 (C9), 51.2 (C10), 78.2 (C11), 18.3 (C12), 57.1 (C13), 21.6 (C14), 26.5 (C15), 45.7 (C16), 53.1 (C17), 18.3 (C18), 61.9 (C19), 170.0 (C20), 21.2 (C21).

Crystal data

C ₂₁ H ₂₄ N ₂ O ₃	<i>Z</i> = 4
<i>M_r</i> = 352.42	<i>D_x</i> = 1.307 Mg m ⁻³
Orthorhombic, <i>P</i> 2 ₁ 2 ₁	Cu Kα radiation
<i>a</i> = 9.436 (1) Å	<i>μ</i> = 0.71 mm ⁻¹
<i>b</i> = 10.588 (1) Å	<i>T</i> = 297 (2) K
<i>c</i> = 17.926 (2) Å	Block, brown
<i>V</i> = 1791.0 (3) Å ³	0.18 × 0.15 × 0.08 mm

Data collection

Enraf–Nonius CAD-4 diffractometer	1135 reflections with <i>I</i> > 2σ(<i>I</i>)
<i>ω</i> / <i>θ</i> scans	<i>R</i> _{int} = 0.072
Absorption correction: none	<i>θ</i> _{max} = 68.0°
3454 measured reflections	2 standard reflections
1788 independent reflections	frequency: 120 min
	intensity decay: 3%

Refinement

Refinement on F^2	$w = 1/[\sigma^2(F_o^2) + (0.053P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.040$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.107$	$(\Delta/\sigma)_{\max} < 0.001$
$S = 1.01$	$\Delta\rho_{\max} = 0.19 \text{ e } \text{\AA}^{-3}$
1788 reflections	$\Delta\rho_{\min} = -0.19 \text{ e } \text{\AA}^{-3}$
236 parameters	Extinction correction: <i>SHELXL97</i>
H-atom parameters constrained	(Sheldrick, 1997)
	Extinction coefficient: 0.0016 (3)

Table 1

Selected geometric parameters (\AA , $^\circ$).

O1—C19	1.405 (5)	C7—C13	1.493 (5)
O2—C12	1.454 (4)	C9—C10	1.517 (5)
O3—C20	1.207 (5)	C16—C19	1.511 (5)
N1—C7	1.282 (4)	C20—C21	1.469 (6)
N1—C7—C13	125.4 (4)	C13—C7—C8	119.8 (3)
N1—C7—C8	114.8 (3)		
N2—C10—C11—C15	20.2 (4)	N2—C17—C16—C15	22.5 (4)
N2—C13—C14—C15	23.0 (4)		

Table 2

Hydrogen-bond geometry (\AA , $^\circ$).

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
O1—H1 \cdots O3 ¹	0.82	2.03	2.810 (4)	160

Symmetry code: (i) $x + \frac{1}{2}, -y + \frac{3}{2}, -z + 2$.

All H atoms were placed in calculated positions and allowed to ride on their parent atoms, with C—H distances in the range 0.93–0.98 \AA and an O—H distance of 0.82 \AA , and with $U_{\text{iso}}(\text{H}) = 1.5U_{\text{eq}}(\text{C})$ for methyl H atoms or $1.2U_{\text{eq}}(\text{C}, \text{O})$ for all other atoms. Shorter than expected bonds and high displacement components along these bonds are found for the terminal methyl C atoms and the indole ring (Spek, 2003; Hirshfeld, 1976).

Due to poor data quality and the absence of any significant anomalous scatterers in the compound, the absolute configuration could not be determined reliably and the 1279 Friedel pairs were

merged before the final refinement. The present configuration is that defined in *Chemical Abstracts* (CAS No. 36285–11-7).

Data collection: *CAD-4 Software* (Enraf–Nonius, 1994); cell refinement: *CAD-4 Software*; data reduction: *XCAD4* (Harms & Wocadlo, 1995); program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEP-3 for Windows* (Farrugia, 1997); software used to prepare material for publication: *WinGX* (Farrugia, 1999).

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References

- Allen, F. H. (2002). *Acta Cryst.* **B58**, 380–388.
- Court, W. E. (1983). *Planta Med.* **48**, 228–233.
- Cremer, D. & Pople, J. A. (1975). *J. Am. Chem. Soc.* **97**, 1354–1358.
- Djerassi, C., Monteiro, H. J., Walsler, A. & Durham, L. J. (1966). *J. Am. Chem. Soc.* **88**, 1792–1798.
- Enraf–Nonius (1994). *CAD-4 Software*. Version 5.0. Enraf–Nonius, Delft, The Netherlands.
- Farrugia, L. J. (1997). *J. Appl. Cryst.* **30**, 565.
- Farrugia, L. J. (1999). *J. Appl. Cryst.* **32**, 837–838.
- Harms, K. & Wocadlo, S. (1995). *XCAD4*. University of Marburg, Germany.
- Hirshfeld, F. L. (1976). *Acta Cryst.* **A32**, 239–244.
- Kato, L., Braga, R. M., Koch, I. & Kinoshita, L. S. (2002). *Phytochemistry*, **60**, 315–320.
- Libot, F., Kunesch, N. & Poisson, J. E. (1980). *Phytochemistry*, **19**, 989–991.
- Libot, F., Miet, C., Kunesch, N., Poisson, J. E., Pusset, J. & Sevenet, T. (1986). *Ann. Pharm. Fr.* **44**, 477–485.
- Li, J. & Cook, J. M. (1998). *J. Org. Chem.* **63**, 4166–4167.
- Prewo, R. & Stezowski, J. J. (1978). *Acta Cryst.* **B34**, 454–460.
- Sheldrick, G. M. (1997). *SHELXS97* and *SHELXL97*. University of Göttingen, Germany.
- Siddiqui, S., Haider, S. I., Ahmad, S. S. & Sultana, N. (1987). *Pak. J. Sci. Ind. Res.* **30**, 71–80.
- Solis, P. N., Wright, C. W., Gupta, M. P. & Phillipson, J. D. (1993). *Phytochemistry*, **33**, 1117–1119.
- Spek, A. L. (2003). *J. Appl. Cryst.* **36**, 7–13.
- Stöckigt, J. (1995). *The Alkaloids: Chemistry and Pharmacology*, edited by G. A. Cordell, pp. 115–172. San Diego: Academic Press.